

# Test Order

## GeneXpert (MTB/RIF)

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<b>Performing Laboratory</b>	Santa Barbara County Public Health Laboratory
<b>Specimen Type</b>	Varies. Please see <i>Mycobacteriology (AFB culture and smear)</i> requirements for a comprehensive list of sample options.
<b>Specimen Container</b>	Sterile collection container (100 ml Falcon Tube preferred)
<b>Minimum Volume</b>	Sterile samples: 1 ml; nonsterile samples: 0.5 ml.
<b>Specimen Handling (Collection Information)</b>	<p><b>When this test is ordered, reflex tests may be performed and charged.</b> The GeneXpert is a reflex test associated with the AFB culture and smear. The assay may be requested by providers, but it is never run independently of the AFB culture and smear. Positive smears will automatically be reflexed to include a GeneXpert. For a more comprehensive list of sample sources and collection procedures, consult the test <i>Mycobacteriology (AFB culture and smear)</i>.</p> <p><b>Sterile samples being analyzed using this methodology, including the procedures for the AFB culture and smear, require a minimum volume of 2.5 ml. For non-sterile samples, both procedures require a minimum volume of 3 mls.</b> The latter sample volumes are the minimum requirements needed for primary and reflex testing methodologies. These minimum volumes will not permit retesting.</p> <p>At this time, the Santa Barbara Public Health Laboratory is unable to analyze samples from pediatric patients (ages 0-17) without qualification.</p>
<b>Transportation Information</b>	Refrigerate at 2-8°C and transport to the laboratory as soon as possible. For more detailed source dependent transportation information, consult the test <i>Mycobacteriology (AFB culture and smear)</i> .
<b>Specimen Stability</b>	Up to seven days refrigerated.
<b>Unacceptable Conditions</b>	Samples that have exceeded the seven day holding period, specimens contaminated/diluted with other liquid substances, specimens transported in gel/anaerobic collection containers, frozen samples/samples transported on dry ice, and specimens that are received unlabeled/mislabeled and/or without the appropriate Santa Barbara Public Health Laboratory requisition
<b>CPT Codes</b>	87556

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# Test Order

Mycobacteriology (AFB culture and smear)

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<b>Performing Laboratory</b>	Santa Barbara County Public Health Laboratory
<b>Specimen Type</b>	<ol style="list-style-type: none"><li>1. Abscesses and Wounds</li><li>2. Blood</li><li>3. Body Fluids</li><li>4. Bone Marrow</li><li>5. Cerebrospinal Fluid (CSF)</li><li>6. Gastric Lavage</li><li>7. Tissue</li><li>8. Respiratory specimens (Bronchoalveolar lavage fluids, bronchial washing, sputum, tracheal aspirates)</li><li>9. Stool</li><li>10. Urine</li></ol>
<b>Specimen Container</b>	Sterile container (100 ml Falcon Tube preferred), evacuated container, syringe.
<b>Minimum Volume</b>	2.5 ml
<b>Specimen Handling (Collection Information)</b>	<p><b>When this test is ordered, reflex tests may be performed and charged.</b></p> <p><u>Abscesses and Wounds</u> Collect sample using a culture transport swab (noncharcoal) Culturette. <b>Samples collected in charcoal or gel based media will not be accepted by the laboratory.</b> Before collecting specimen, wipe away excessive secretions and/or discharge. Obtain secretions or fluid from area of interest with sterile swab or via aspiration. If smear and culture are requested or if both a bacterial culture and mycobacterial culture are requested from the same collection location, collect an additional swab to maximize test sensitivity. For low volume samples, swab the area of interest and insert the swab into a sterile collection container with 2 ml of sterile DI water or 0.5% saline.</p> <p><u>Blood</u> Draw sample directly into a <i>Bactec Myco/F Lytic</i> blood culture bottle following all established venipuncture procedures to avoid cross contamination. <i>Myco/F Lytic</i> blood culture bottles can be provided to clinics upon request.</p> <p><u>Body Fluids</u> Obtain sample from area of interest and transfer into sterile, approved collection container.</p> <p><u>Bone Marrow</u> Obtain sample from area of interest and transfer into sterile, approved collection container. Do not mix samples to be tested for AFB with preservatives that may be necessary for other requested assays (i.e., Vital Transport Media (VTM)). Samples may be collected with</p>

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# Test Order

## Mycobacteriology (AFB culture and smear)

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containers containing lithium heparin, but unpreserved specimens are preferred.

### CSF

Perform a lumbar puncture collecting samples into pre-labeled CSF tubes (e.g., tubes one through four). Aliquot 1-2 ml into sterile collection container or send the contents of tube three to the Public Health Laboratory.

### Gastric Lavage

Following approved procedures, administer and aspirate a volume of fluid via an orogastric tube. Collect the contents in a sterile collection container and **neutralize the gastric sample within four hours using 100 mg of sodium bicarbonate per 5 to 10 mL of gastric wash.** Specimens should be tested using a pH strip prior to transportation to the laboratory. **An acceptable sample will be neutralized to a pH of 7.0-7.5.** Samples that arrive at the laboratory that are either excessively neutralized or inappropriately neutralized (i.e., pH 6.2) will not be tested.

### Tissue

Excise a fresh tissue sample from area of interest. When applicable, excise portions of necrotic tissue for analysis. Homogenize tissue with sterile DI water or 0.5 % saline and transfer into Falcon tube. **If homogenization prior to delivery to the laboratory is not possible, add 1-2 ml of sterile DI water or 0.5 % saline to prevent sample from desiccating.** Samples that are not homogenized prior to delivery will be homogenized by the laboratory at an additional cost.

### Respiratory specimens

Collect three respiratory specimens for patients with clinical and chest X-ray findings indicating tuberculosis. **These three specimens must be collected at 8 to 24 hour intervals (when possible, a 24 hour interval is preferred).** At least one of the three samples should include a first morning specimen.

### Stool

Obtain a random stool sample and transfer contents into sterile collection container.

### Urine

Obtain a random urine specimen and transfer sample into sterile collection container.

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### **Transportation**

For all samples **except** those collected into the *Bactec Myco/F Lytic*

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## Test Order

### Mycobacteriology (AFB culture and smear)

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<b>Information</b>	<p>culture vial: refrigerate at 2-8°C and transport to the laboratory as soon as possible.</p> <p>For samples collected using the <i>Bactec Myco/F Lytic</i> culture vial: transport specimens at ambient temperatures. Deliver to the laboratory as soon as possible without exceeding 72 hours</p>
<b>Specimen Stability</b>	<p>For all samples <b>except</b> those collected into the <i>Bactec Myco/F Lytic</i> culture vial: up to seven days refrigerated.</p> <p>For samples collected using the <i>Bactec Myco/F Lytic</i> culture vial: Up to 72 hours.</p>
<b>Unacceptable Conditions</b>	<p>Specimens received leaking or in a broken transport container, samples collected in gel or charcoal based transport media, specimens that have surpassed the seven day testing window, samples received in blood culture bottles other than those approved for the recovery of mycobacteria, Myco/F Lytic culture bottles that have exceeded 72 hours since the time of collection, and samples that are received unlabeled/mislabeled and/or without the appropriate Santa Barbara Public Health Laboratory requisition.</p>
<b>CPT Codes</b>	87206, 87015, 87116

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